

---

## Soil Microbial Processes and the Cycling of Atmospheric Trace Gases [and Discussion]

Ralf Conrad and K. A. Smith

*Phil. Trans. R. Soc. Lond. A* 1995 **351**, 219-230

doi: 10.1098/rsta.1995.0030

---

### Email alerting service

Receive free email alerts when new articles cite this article - sign up in the box at the top right-hand corner of the article or click [here](#)

---

To subscribe to *Phil. Trans. R. Soc. Lond. A* go to:

<http://rsta.royalsocietypublishing.org/subscriptions>

---

# Soil microbial processes and the cycling of atmospheric trace gases

BY RALF CONRAD

*Max-Planck-Institut für Terrestrische Mikrobiologie, Karl-von-Frisch-Str.,  
D-35043 Marburg, Germany*

Soil microbial processes involved in the flux between soil and atmosphere of the atmospheric trace gases methane ( $\text{CH}_4$ ), hydrogen ( $\text{H}_2$ ), carbon monoxide ( $\text{CO}$ ), carbonyl sulphide ( $\text{OCS}$ ), nitrous oxide ( $\text{N}_2\text{O}$ ), and nitric oxide ( $\text{NO}$ ) are reviewed. The flux of a trace gas between soil and atmosphere is usually the result of simultaneous production and consumption reactions, so that a compensation concentration exists, at which the net flux is zero. With some of the trace gases, different suites of processes operate at different gas concentrations, so that the processes that consume a trace gas that is produced within the soil are often different from those that consume the trace gas entering the soil from the atmosphere. Certain groups of soil microorganisms can metabolize more than one of these trace gases. The processes involved in the cycling of a trace gas in anoxic wetland soils are often different from those operating in upland soils.

## 1. Introduction

Junge (1972) pointed out that virtually all the atmospheric gases undergo cycles which are more or less dominated by the biosphere. Only a few species (e.g.  $\text{O}_3$ , Rn) exhibit cycles which are purely governed by physical-chemical reactions. The main sources and sinks for atmospheric trace gases in the biosphere are the ocean, soils, vegetation, and animals: for detailed reviews see Buat-Menard (1986), Bouwman (1990), Sharkey *et al.* (1991), and Khalil (1993).

The sources and sinks in soils include (1) abiotic chemical reactions (e.g. the production of  $\text{CO}$  from soil organic carbon (Conrad & Seiler 1985*a*)), (2) 'fortuitous' biochemical reactions that do not support microbial growth (e.g. hydrolysis of  $\text{OCS}$  (Kesselmeier 1992)), (3) biochemical reactions by the extracellular enzymes present in soil (e.g. oxidation of  $\text{H}_2$  (Conrad *et al.* 1983*a*)), and (4) metabolic reactions that support the growth of microorganisms (e.g. oxidation of  $\text{CH}_4$  (King 1992)).

In most of these processes microorganisms are either directly or indirectly involved. Soils probably show the highest abundance and diversity of microorganisms in nature, and thus it is no wonder that soils are involved in and can even dominate the cycling of many atmospheric trace gases. Table 1 lists a selection of atmospheric trace gases for which soils are important sources and/or sinks, and which are all (with the possible exception of  $\text{H}_2$ ) of importance in atmospheric chemistry and thus of considerable concern with respect to climate and to global change. To understand and predict changes in the source and sink strengths of

Table 1. *Contribution of the soil to the cycling of atmospheric trace gases*

trace gas	lifetime (days)	mixing ratio (ppbv)	total budget (Tg a <sup>-1</sup> )	annual increase (%)	contribution (%) of soils to total budgets as		impact	ref.
					source	sink		
N <sub>2</sub> O	60 000	310	15	0.2–0.3	70	?	stratospheric chemistry; greenhouse effect	1–4
CH <sub>4</sub>	4000	1700	540	< 0.8	60	5	greenhouse effect; tropospheric & stratospheric chemistry	4–7
H <sub>2</sub>	1000	550	90	0.6	5	95	insignificant	8,9
OCS	> 350	0.5	1.2	?	25	?	aerosol formation	10–12
CO	100	100	2600	1.0 <sup>a</sup>	1	15	tropospheric chemistry	8,13
NO	1	< 0.1	60	?	20	?	tropospheric chemistry	3,14

(1) Khalil & Rasmussen 1992; (2) Bouwman 1990; (3) Davidson 1991; (4) Prinn 1994; (5) Cicerone & Oremland 1988; (6) Khalil & Rasmussen 1990a; (7) Steele *et al.* 1992; (8) Conrad 1988; (9) Khalil & Rasmussen 1990b; (10) Andreae & Jaeschke 1992; (11) Chin & Davis 1993; (12) Möller 1984; (13) Khalil & Rasmussen 1990c; (14) Conrad 1990.

<sup>a</sup>The long-term trend has recently reversed; CO is now decreasing by about 6% per year (Novelli *et al.* 1994).

these trace gases, it is necessary to know the processes involved in generating the net flux at the soil–atmosphere interface.

## 2. Diffusion of trace gases

The net flux of a trace gas is usually the result of production, consumption, and transport within the soil column. Whether produced or consumed, the diffusional flux of a trace gas is basically controlled by Fick's law and thus is proportional to the actual diffusion coefficient in soil and to the concentration gradient of the trace gas. The actual diffusion coefficient is difficult to determine, being affected by the temperature and air-filled porosity of the soil. In particular, the distribution of water has drastic effects on diffusion. In well-aerated soils ('upland soils'), the diffusional flux is dominated by diffusion in the air-filled soil pores, since the molecular diffusion coefficient in water is about four orders of magnitude lower than in air. In addition, many of the trace gases (in table 1 all except N<sub>2</sub>O) are not very soluble in water. However, in watersaturated flooded soils ('wetland soils'), diffusion is entirely within the water phase until the trace gas reaches the water–atmosphere interface or a plant root, from where it may be transported to the atmosphere by gas mass flow or by diffusion through the plant's gas vascular system (Armstrong 1979; Schütz *et al.* 1991). Some of the differences between upland and wetland soils are depicted in figure 1.

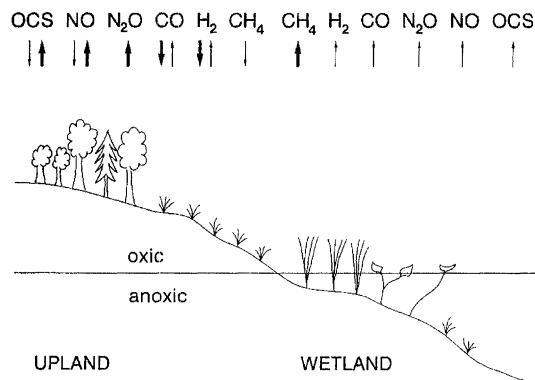


Figure 1. Flux of trace gases between upland and wetland soils and the atmosphere. Thick arrows represent major fluxes, thin arrows minor fluxes.

### 3. Simultaneous production and consumption of trace gases in soil

Two different situations exist. The first is where production and consumption processes are more or less homogeneously distributed within the soil, a situation typical of upland soils, but also of anaerobic processes occurring within the anoxic zones of wetland soils. The second is where production and consumption processes are spatially separated, as is usual in wetland soils, where the deeper anoxic soil layers (sites of production) are covered at the surface with a shallow oxic layer (site of consumption). The roots of aquatic macrophytes that penetrate the anoxic zones of wetland soils are often surrounded by a thin oxic layer, which also provides a site for aerobic consumption processes (Conrad 1989; Schütz *et al.* 1991).

In upland soils, the existence of both production and consumption reactions can be inferred if it can be shown that there is a compensation concentration at which the two processes are in equilibrium and at which no net flux is observed. In oxic upland soils this is almost always true, with compensation concentrations established for  $H_2$  (Seiler 1978; Conrad & Seiler 1980a),  $CO$  (Seiler 1978; Conrad & Seiler 1985b),  $OCS$ ,  $N_2O$  (Seiler & Conrad 1981) and  $NO$  (Johansson & Galbally 1984; Remde *et al.* 1989). The compensation concentrations for  $N_2O$  are usually so high that upland soils are almost always a source of atmospheric  $N_2O$ . This may also turn out to be the case for  $OCS$ . By contrast, the compensation concentrations for  $H_2$  are usually so low (except at sites with biological  $N_2$  fixation (Conrad & Seiler 1980a)), that upland soils almost always act as a sink for atmospheric  $H_2$  (Seiler 1978; Conrad 1988). On the other hand, the compensation concentrations for  $CO$  (Conrad & Seiler 1980b, 1985b; Scharffe *et al.* 1990) and  $NO$  (Slemr & Seiler 1984, 1991) vary with changing soil conditions and are in a range such that soil may dynamically change from acting as a source to acting as a sink. An example is shown in figure 2, where the  $CO$  compensation concentration changes during the day. During the hot hours of the day it is higher than the atmospheric  $CO$  concentration, so that there is a net flux of  $CO$  from the soil into the atmosphere. If the  $CO$  concentration is increased experimentally,  $CO$  is taken up by the soil until the compensation concentration is once again reached.

There is, as yet, no firm evidence for a  $CH_4$  compensation concentration. However, a compensation concentration may exist, since tropical upland soils are

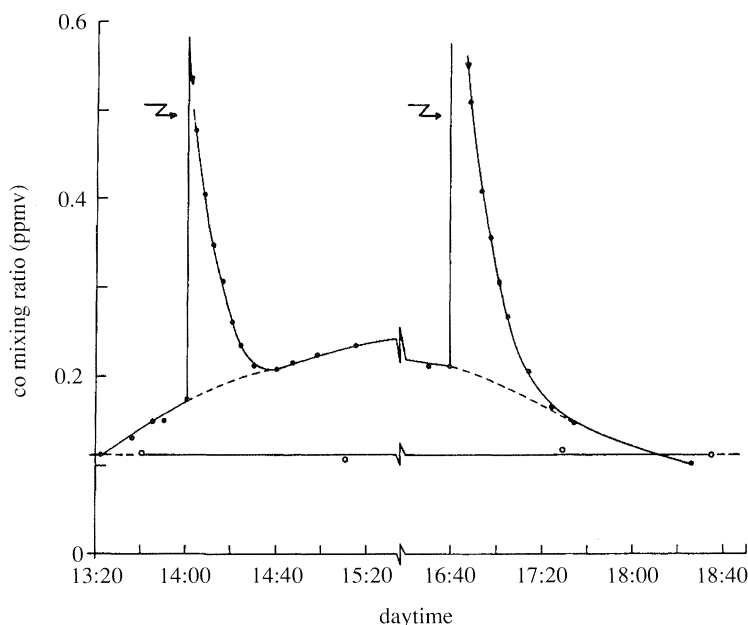


Figure 2. Diurnal change of the CO concentration outside (o) and inside (•) a chamber covering the surface of a savanna soil in the Transvaal (South Africa). The CO concentrations inside the chamber represent CO compensation concentrations between release and uptake by the soil. The arrows indicate injection of CO into the chamber. Adapted from Conrad and Seiler (1985c).

sometimes sources of atmospheric  $\text{CH}_4$ , especially during the rainy season (Keller *et al.* 1986, 1993). However, in most upland soils  $\text{CH}_4$  production seems to be marginal, probably because  $\text{CH}_4$ -producing bacteria require strictly anoxic and reduced conditions and thus are inactive in well-aerated upland soils (Cicerone & Oremland 1988; Conrad 1989; Reeburgh *et al.* 1993). Methane uptake by upland soils seems to be limited by diffusion from the atmosphere into the soil (Dörr *et al.* 1993) and from the air-filled pore space into the soil aggregates (Koschorreck & Conrad 1993). It is also limited by the activity of methanotrophic bacteria, which typically occur in subsurface soil layers and on the larger soil grains (Bender & Conrad 1994), and by the threshold and affinity for  $\text{CH}_4$  of the methanotrophic population (see below; Bender & Conrad 1992, 1993).

In wetland soils, oxidation of  $\text{CH}_4$  in the overlying oxic soil layer diminishes the flux of  $\text{CH}_4$  to the atmosphere from the deeper water-saturated, anoxic soil (Reeburgh *et al.* 1993). It is also possible that  $\text{CH}_4$  is oxidized during its passage through the oxic layers surrounding the roots of these plants that penetrate the anoxic layers. The role of transport and oxidation in the cycling of  $\text{CH}_4$  in paddy fields has recently been reviewed by Conrad (1993).

#### 4. Gas concentration and microbial activity

Microorganisms metabolizing trace gases at atmospheric concentrations must be able to deal with gas concentrations that are typically in the picomolar to nanomolar range. These low concentrations are unfavourable for synthesis of cellular biomass (Conrad 1984). Interestingly, many of the processes involved in the consumption of trace gases at atmospheric concentrations are fortuitous reac-

Table 2. *Microorganisms and reactions important in production or consumption of trace gases in soil*

gas	trace production	consumption at elevated concentrations	consumption at atmospheric concentrations
CH <sub>4</sub>	methanogens	common methanotrophs; nitrifiers	unknown methanotrophs; nitrifiers
H <sub>2</sub>	fermenting bacteria; N <sub>2</sub> -fixing bacteria	Knallgas bacteria; anaerobic chemolithotrophs <sup>a</sup>	abiotic soil hydrogenases
CO	chemical conversion of soil organic carbon; anaerobic chemolithotrophs	carboxydrotrophs; anaerobic chemolithotrophs	nitrifiers; oligotrophic bacteria
OCS	thiocyanate hydrolase	carbonic anhydrase	carbonic anhydrase
N <sub>2</sub> O	denitrifiers; DNRA <sup>b</sup> ;	denitrifiers	denitrifiers
NO	denitrifiers; DNRA; nitrifiers; chemical decomposition of NO <sub>2</sub> <sup>-</sup>	denitrifiers; methanotrophs; heterotrophic bacteria	denitrifiers; methanotrophs; heterotrophic bacteria

<sup>a</sup>Anaerobic chemolithotrophs are methanogens, homoacetogens, sulphate reducers, etc.

<sup>b</sup>DNRA is dissimilatory reduction of nitrate to ammonium.

tions that do not serve microorganisms for growth (table 2). This is the case with the oxidation of CO by ammonium-oxidizing nitrifiers (Jones & Morita 1983a; Jones *et al.* 1984), of H<sub>2</sub> by soil hydrogenases (Conrad & Seiler 1981; Conrad *et al.* 1983a; Schuler & Conrad 1990), of OCS by the carbonic anhydrase present in various CO<sub>2</sub>-fixing bacteria (Protoschill-Krebs & Kesselmeier 1992), and of NO by heterotrophic bacteria (Baumgärtner *et al.* 1995, unpublished work). In all these cases, the trace gases are oxidized rather than reduced and the enzymes involved have the high affinity and the low threshold required for reaction with a trace gas at atmospheric concentrations. This is also true for atmospheric CH<sub>4</sub> which, however, is probably not fortuitously but 'usefully' oxidized by an unknown methanotrophic population (Bender & Conrad 1992). Similarly, atmospheric CO seems to be oxidized to some extent by unknown oligotrophic microorganisms (Conrad & Seiler 1982).

Only N<sub>2</sub>O and (to some extent) NO seem to be consumed by reduction processes in aerated upland soils. Such soils normally contain anoxic microsites (Sexton *et al.* 1985; Zausig *et al.* 1993) which are presumably where the reduction occurs. Reduction of NO and N<sub>2</sub>O mainly involves the well-known denitrifiers that catalyse the sequential reduction of nitrate → nitrite → NO → N<sub>2</sub>O → N<sub>2</sub> (Knowles 1982; Tiedje 1988; Zafiriou *et al.* 1989; Remde & Conrad 1991). However, consumption of atmospheric NO in upland soils seems to involve two additional reactions, (i) fortuitous oxidation by heterotrophic bacteria (see above) and (ii) consumption by methanotrophic bacteria (Krämer *et al.* 1990).

Frequently, however, the microorganisms involved in the consumption of trace gases at atmospheric concentrations are not those which have been isolated on the basis of their ability to use one of the trace gases for growth at high gas concentrations (percent level). This is obviously true for the commonly known methanotrophs (CH<sub>4</sub>) (Conrad 1984; Bender & Conrad 1992; King 1993), the Knallgas bacteria (H<sub>2</sub>) (Conrad 1988), and the carboxydrotrophic bacteria (CO)

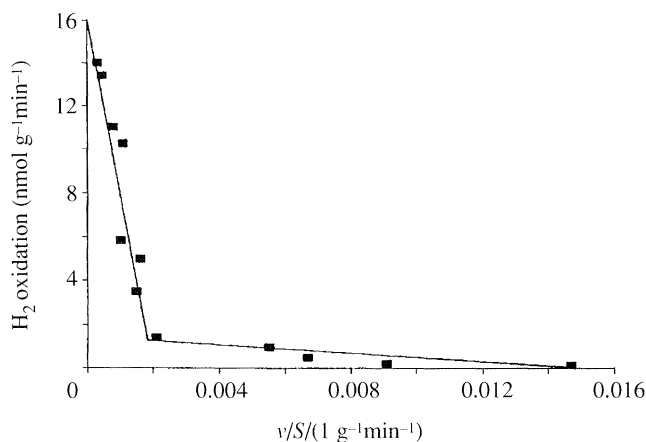


Figure 3. Eadie-Hofstee plot of the kinetics of  $H_2$  oxidation in a meadow soil, revealing two activities with  $K_m$  values of 19 and 1450 ppmv  $H_2$ , respectively. Adapted from Häring & Conrad (1994).

(Conrad 1988). All these bacteria can only use trace gases at elevated concentrations (table 2). For this reason they are probably only active adjacent to special spots (e.g. anoxic microsites, root nodules; see below) in the soil where the trace gases are produced at such high rates that elevated concentrations are generated. Thus, Knallgas bacteria contribute to the oxidation of the relatively high  $H_2$  concentrations found in the vicinity of  $H_2$ -producing root nodules of legumes (LaFavre & Focht 1983; Popelier *et al.* 1985; Cunningham *et al.* 1986; Schuler & Conrad 1991). Kinetic experiments demonstrate the existence in soil of both Knallgas bacteria, with low affinity for  $H_2$  and soil hydrogenases, with high affinity for  $H_2$  (figure 3; Schuler & Conrad 1990; Häring & Conrad 1994). Schuler & Conrad (1991) showed that the  $H_2$  compensation concentration was relatively large when nodulated legumes grown on sterile soil were inoculated with Knallgas bacteria. The Knallgas bacteria could only consume the  $H_2$  produced by the legumes down to concentrations of about 13 ppmv  $H_2$ . The consumption of  $H_2$  to still lower concentrations was only possible if the soil hydrogenases were also active, i.e. in non-sterile soil. However, it is likely that the soil hydrogenases operate in tandem with the  $H_2$ -oxidizing bacteria, since the soil hydrogenases are reversibly inhibited by exposure to high  $H_2$  concentrations (Conrad & Seiler 1981). Knallgas bacteria presumably first reduce the  $H_2$  concentration to a level that allows the soil hydrogenases to operate.

The importance of these microorganisms for reducing the flux of trace gases into the atmosphere is most evident in wetland soils (e.g. rice fields), where the metabolites  $CH_4$ ,  $H_2$ ,  $N_2O$  and  $NO$  are produced at potentially high rates but released into the atmosphere at drastically reduced rates. Consumption of  $H_2$  is achieved by various anaerobic chemolithotrophic bacteria (methanogens, homoacetogens, sulphate reducers, etc.) within the anoxic environment (Conrad 1989) and, in addition, by the aerobic Knallgas bacteria at the anoxic-oxic interface (Schütz *et al.* 1988). Consumption of  $CH_4$  is achieved by the common methanotrophic bacteria at the anoxic-oxic interface (King 1990; Conrad & Rothfuss 1991; Bosse *et al.* 1993; Bender & Conrad 1994). Nitrous oxide and  $NO$  are effectively consumed within the anoxic environment by sequential reductive den-

itrification to  $N_2$ , thus reducing the flux into the atmosphere to very low rates (Freney & Denmead 1992; Galbally *et al.* 1987), but in this case by the same denitrifying bacteria which act on NO and  $N_2O$  at atmospheric concentrations.

### 5. Microorganisms producing or consuming more than one trace gas

Certain species of microorganism are capable of metabolizing more than one trace gas; see table 2. Autotrophic nitrifiers are involved in the oxidation of CO (Jones & Morita 1983*a*; Jones *et al.* 1984) and probably also of  $CH_4$  (Hyman & Wood 1983; Jones & Morita 1983*b*; Castro *et al.* 1994). Methanotrophs can also be involved in heterotrophic nitrification and thus in the cycling of NO and  $N_2O$  (Yoshinari 1985; Knowles & Topp 1988; Krämer *et al.* 1990). The significance of the bioversatility of these organisms for the turnover of trace gases in soil deserves further investigation. Several types of anaerobic  $H_2$ -consuming chemolithotrophic bacteria (methanogens, homoacetogens, sulphate reducers, etc.), which play an important role in the control of  $CH_4$  production in anoxic wetland soils (Conrad 1989), also have the potential to produce and consume CO (Diekert & Wohlfarth 1994). However, the significance of this potential for the turnover of CO in wetland soils is largely unexplored (Conrad *et al.* 1983*b*, 1988).

### 6. Cycling of trace gases in wetland and upland soils

Whereas wetland soils are always sources, upland soils can act both as sources or sinks of atmospheric trace gases (figure 1). For the most part, we do not know whether the microorganisms living in anoxic microsites within upland soils are the same as those living in anoxic wetland soils. Interestingly, strictly anoxic methanogenic bacteria that typically occur in anoxic wetland soils also occur in low numbers in upland soils which have never been flooded (Mayer & Conrad 1990); they are even found in desert soils (Peters & Conrad 1995). As a first approach, we may therefore assume that although the species might be different, the reactions that they catalyse are the same in both environments.

However, anaerobic microorganisms are both more abundant and more active in wetland than in upland soils. This is obvious for production processes such as  $CH_4$  production (Conrad 1989). In principle, it is also true for  $H_2$  which is produced in large amounts by fermenting bacteria in wetland soils, but converted to  $CH_4$  (among other products) so that only small amounts of  $H_2$  escape into the atmosphere (Schütz *et al.* 1988). In upland soils, on the other hand,  $H_2$  production by fermentation is negligible (Conrad 1988). Instead, it is produced by biological  $N_2$  fixation in legumes (Conrad & Seiler 1980*a*), whose root nodules provide a highly specialized low- $O_2$  environment, allowing the  $O_2$ -sensitive nitrogenase to be active within oxic soils.

Very little is known about how the production of CO and OCS differ in upland and wetland soils. In oxic upland soils, CO is mainly produced during the chemical oxidation of soil organic matter (Conrad & Seiler 1985*a, b*). In anoxic wetland soils, CO can also be produced by anaerobic chemolithotrophs during metabolism (Diekert & Wohlfarth 1994). The production processes of OCS are mostly unknown, except for one reaction, the hydrolysis of thiocyanate ( $SCN^- + H_3O^+ \rightarrow OCS + NH_3$ ) by microorganisms containing the enzyme thiocyanate



hydrolase (Kelly *et al.* 1993; Katayama *et al.* 1992). This enzyme may be active in both wetland and upland soils (Minami & Fukushi 1981).

The difference between upland and wetland soils is also seen in consumption processes, for example H<sub>2</sub> consumption. In contrast to upland soils, where soil hydrogenases and, to some extent, also Knallgas bacteria collaborate (Conrad 1988), consumption of H<sub>2</sub> in wetland soils is achieved by anaerobic chemolithotrophs (Conrad 1989, 1995). A similar difference exists for CO consumption. Ammonium-oxidizing nitrifiers, oligotrophic bacteria, and (to some extent) carboxydrotrophic bacteria all contribute to CO consumption in upland soils (Conrad 1988; Mörsdorf *et al.* 1992), whereas it is likely that anaerobic chemolithotrophs dominate CO turnover in wetland soils. The methanotrophs active in wetland soils differ in their kinetics from those in upland soils, since the former are exposed to elevated CH<sub>4</sub> concentrations, whereas the latter are exposed to atmospheric concentrations of CH<sub>4</sub> (see above; Bender & Conrad 1992, 1994). Anaerobic CH<sub>4</sub>-oxidizing microorganisms seem to play an important role in marine sediments (Alperin & Reeburgh 1984; Iversen & Joergensen 1985), but have not yet been isolated.

### References

- Alperin, M. J. & Reeburgh, W. S. 1984 Geochemical observations supporting anaerobic methane oxidation. In *Microbial growth on C-1 compounds* (ed. R. L. Crawford & R. S. Hanson), pp. 282–289. Washington, D.C.: American Society for Microbiology.
- Andreae, M. O. & Jaeschke, W. A. 1992 Exchange of sulphur between biosphere and atmosphere over temperate and tropical regions. In *Sulphur cycling on the continents* (ed. R. W. Howarth, J. W. B. Stewart & M. V. Ivanov), *SCOPE Rep.* **48**, 27–61. Wiley.
- Armstrong, W. 1979 Aeration in higher plants. *Adv. bot. Res.* **7**, 226–332.
- Bender, M. & Conrad, R. 1992 Kinetics of CH<sub>4</sub> oxidation in oxic soils exposed to ambient air or high CH<sub>4</sub> mixing ratios. *FEMS Microbiol. Ecol.* **101**, 261–270.
- Bender, M. & Conrad, R. 1993 Kinetics of methane oxidation in oxic soils. *Chemosphere* **26**, 687–696.
- Bender, M. & Conrad, R. 1994 Methane oxidation activity in various soils and sediments: occurrence, characteristics, vertical profiles and distribution on grain size fractions. *J. geophys. Res.* **99**, 16531–16540.
- Bosse, U., Frenzel, P. & Conrad, R. 1993 Inhibition of methane oxidation by ammonium in the surface layer of a littoral sediment. *FEMS Microbiol. Ecol.* **13**, 123–134.
- Bouwman, A. F. 1990 Exchange of greenhouse gases between terrestrial ecosystems and the atmosphere. In *Soils and the greenhouse effect* (ed. A. F. Bouwman), pp. 61–127. Chichester: Wiley.
- Bouwman, A. F. (ed.) 1990 *Soils and the greenhouse effect*. Chichester: Wiley.
- Buat-Menard, P. (ed.) 1986 *The role of air–sea exchange in geochemical cycling*. Dordrecht: Reidel.
- Castro, M. S., Peterjohn, W. T., Melillo, J. M., Steudler, P. A., Gholz, H. L. & D. Lewis 1994 Effects of nitrogen fertilization on the fluxes of N<sub>2</sub>O, CH<sub>4</sub> and CO<sub>2</sub> from soils in a Florida slash pine plantation. *Can. J. Forest Res.* **24**, 9–13.
- Chin, M. & Davies, D. D. 1993 Global sources and sinks of OCS and CS<sub>2</sub> and their distributions. *Global Biogeochem. Cycles* **7**, 321–337.
- Cicerone, R. J. & Oremland, R. S. 1988 Biogeochemical aspects of atmospheric methane. *Global Biogeochem. Cycles* **2**, 299–327.
- Conrad, R. 1984 Capacity of aerobic microorganisms to utilize and grow on atmospheric trace gases (H<sub>2</sub>, CO, CH<sub>4</sub>). In *Current perspectives in microbial ecology* (ed. M. G. Klug & C. A. Reddy), pp. 461–467. Washington, DC: American Society for Microbiology.

- Conrad, R. 1988 Biogeochemistry and ecophysiology of atmospheric CO and H<sub>2</sub>. *Adv. Microb. Ecol.* **10**, 231–283.
- Conrad, R. 1989 Control of methane production in terrestrial ecosystems. In *Exchange of trace gases between terrestrial ecosystems and the atmosphere, Dahlem Konferenzen* (ed. M. O. Andreae & D. S. Schimel), pp. 39–58. Chichester: Wiley.
- Conrad, R. 1990 Flux of NO<sub>x</sub> between soil and atmosphere: importance and soil microbial metabolism. In *Denitrification in soil and sediment* (ed. N. P. Revsbech & J. Soerensen), pp. 105–128. New York: Plenum.
- Conrad, R. 1993 Mechanisms controlling methane emission from wetland rice fields. In *The biogeochemistry of global change: radiative trace gases* (ed. R. S. Oremland), pp. 317–335. New York: Chapman & Hall.
- Conrad, R. 1995 Anaerobic hydrogen metabolism in aquatic sediments. In *Cycling of reduced gases in the hydrosphere* (ed. D. D. Adams, S. P. Seitzinger & P. M. Crill). Stuttgart: Schweitzerbart'sche Verlagsbuchhandlung. (In the press.)
- Conrad, R. & Rothfuss, F. 1991 Methane oxidation in the soil surface layer of a flooded rice field and the effect of ammonium. *Biol. Fertil. Soils* **12**, 28–32.
- Conrad, R. & Seiler, W. 1980a Contribution of hydrogen production by biological nitrogen fixation to the global hydrogen budget. *J. geophys. Res.* **85**, 5493–5498.
- Conrad, R. & Seiler, W. 1980b Role of microorganisms in the consumption and production of atmospheric carbon monoxide by soil. *Appl. Environ. Microbiol.* **40**, 437–445.
- Conrad, R. & Seiler, W. 1981 Decomposition of atmospheric hydrogen by soil microorganisms and soil enzymes. *Soil Biol. Biochem.* **13**, 43–49.
- Conrad, R. & Seiler, W. 1982 Utilization of traces of carbon monoxide by aerobic oligotrophic microorganisms in ocean, lake and soil. *Arch. Microbiol.* **132**, 41–46.
- Conrad, R. & Seiler, W. 1985a Characteristics of abiological CO formation from soil organic matter, humic acids, and phenolic compounds. *Environ. Sci. Technol.* **19**, 1165–1169.
- Conrad, R. & Seiler, W. 1985b Influence of temperature, moisture and organic carbon on the flux of H<sub>2</sub> and CO between soil and atmosphere. Field studies in subtropical regions. *J. geophys. Res.* **90**, 5699–5709.
- Conrad, R. & Seiler, W. 1985c Feldmessung von Emission und Deposition atmosphärischer Spurengase in Boden und Wasser. GIT-Suppl. (Umweltschutz-Umweltanalytik) 3/85, 74–78.
- Conrad, R., Weber, M. & Seiler, W. 1983a Kinetics and electron transport of soil hydrogenases catalyzing the oxidation of atmospheric hydrogen. *Soil Biol. Biochem.* **15**, 167–173.
- Conrad, R., Aragno, M. & Seiler, W. 1983b Production and consumption of carbon monoxide in a eutrophic lake. *Limnol. Oceanogr.* **28**, 42–49.
- Conrad, R., Schütz, H. & Seiler, W. 1988 Emission of carbon monoxide from submerged rice fields into the atmosphere. *Atmos. Environ.* **22**, 821–823.
- Cunningham, S. D., Kapulnik, Y. & Phillips, D. A. 1986 Distribution of hydrogen-metabolizing bacteria in alfalfa field soil. *Appl. Environ. Microbiol.* **52**, 1091–1095.
- Davidson, E. A. 1991 Fluxes of nitrous oxide and nitric oxide from terrestrial ecosystems. In *Microbial production and consumption of greenhouse gases: methane, nitrogen oxides, and halomethanes* (ed. J. E. Rogers & W. B. Whitman), pp. 219–235. Washington, DC: American Society for Microbiology.
- Diekert, G. & Wohlfarth, G. 1994 Kohlenmonoxid im Stoffwechsel strikt anaerober Bakterien. *BioEng.* **10**, 25–32.
- Dörr, H., Katruff, L. & Levin, I. 1993 Soil texture parameterization of the methane uptake in aerated soils. *Chemosphere* **26**, 697–713.
- Freney, J. R. & Denmead, O. T. 1992 Factors controlling ammonia and nitrous oxide emissions from flooded rice fields. *Ecol. Bull. (Copenhagen)* **42**, 188–194.
- Galbally, I. E., Freney, J. R., Muirhead, W. A., Simpson, J. R., Trevitt, A. C. F. & Chalk, P. M. 1987 Emission of nitrogen oxides (NO<sub>x</sub>) from a flooded soil fertilized with urea: relation to other nitrogen loss processes. *J. atmos. Chem.* **5**, 343–365.

- Häring, V. & Conrad, R. 1994 Demonstration of two different H<sub>2</sub>-oxidizing activities in soil using H<sub>2</sub> consumption and a tritium exchange assay. *Biol. Fertil. Soils* **17**, 125–128.
- Hyman, M. R. & Wood, P. M. 1983 Methane oxidation by *Nitrosomonas europaea*. *Biochem. J.* **212**, 31–37.
- Iversen, N. & Joergensen, B. B. 1985 Anaerobic methane oxidation rates at the sulphate–methane transition in marine sediments from Kattegat and Skagerrak (Denmark). *Limnol. Oceanogr.* **30**, 944–955.
- Johansson, C. & Galbally, I. E. 1984 Production of nitric oxide in loam under aerobic and anaerobic conditions. *Appl. Environ. Microbiol.* **47**, 1284–1289.
- Jones, R. D. & Morita, R. Y. 1983a Carbon monoxide oxidation by chemolithotrophic ammonium oxidizers. *Can. J. Microbiol.* **29**, 1545–1551.
- Jones, R. D. & Morita, R. Y. 1983b Methane oxidation by *Nitrosococcus oceanus* and *Nitrosomonas europaea*. *Appl. Environ. Microbiol.* **45**, 401–410.
- Jones, R. D., Morita, R. Y. & Griffiths, R. P. 1984 Method for estimating *in situ* chemolithotrophic ammonium oxidation using carbon monoxide oxidation. *Mar. Ecol. Progr. Ser.* **17**, 259–269.
- Junge, C. 1972 The cycle of atmospheric gases – natural and man made. *Q. Jl R. meteor. Soc.* **98**, 711–729.
- Katayama, Y., Narahara, Y., Inoue, Y., Amano, F., Kanagawa, T. & Kuraishi, H. 1992 A thiocyanate hydrolase of *Thiobacillus thioparus* – a novel enzyme catalyzing the formation of carbonyl sulphide from thiocyanate. *J. Biol. Chem.* **267**, 9170–9175.
- Keller, M., Kaplan, W. A. & Wofsy, S. C. 1986 Emissions of N<sub>2</sub>O, CH<sub>4</sub> and CO<sub>2</sub> from tropical forest soils. *J. geophys. Res.* **91**, 11791–11802.
- Keller, M., Veldkamp, E., Weitz, A. M. & Reiners, W. A. 1993 Effect of pasture age on soil trace-gas emissions from a deforested area of Costa Rica. *Nature, Lond.* **365**, 244–246.
- Kelly, D. P., Malin, G. & Wood, A. P. 1993 Microbial transformations and biogeochemical cycling of one-carbon substrates containing sulphur, nitrogen or halogens. In *Microbial growth on C1 compounds* (ed. J. C. Murrell & D. P. Kelly), pp. 47–63. Andover: Intercept.
- Kesselmeier, J. 1992 Plant physiology and the exchange of trace gases between vegetation and the atmosphere. In *Precipitation scavenging and atmosphere–surface exchange*, vol. 2 (ed. S. E. Schwartz & W. G. N. Slinn), pp. 949–966. Washington: Hemisphere Publishing Corporation.
- Khalil, M. A. K. & Rasmussen, R. A. 1990a Atmospheric methane: recent global trends. *Environ. Sci. Technol.* **24**, 549–553.
- Khalil, M. A. K. & Rasmussen, R. A. 1990b Global increase of atmospheric molecular hydrogen. *Nature, Lond.* **347**, 743–745.
- Khalil, M. A. K. & Rasmussen, R. A. 1990c The global cycle of carbon monoxide: trends and mass balance. *Chemosphere* **20**, 227–242.
- Khalil, M. A. K. & Rasmussen, R. A. 1992 The global sources of nitrous oxide. *J. geophys. Res.* **97**, 14651–14660.
- Khalil, M. A. K. (ed.) 1993 *Atmospheric methane: sources, sinks, and role in global change*. Berlin: Springer.
- King, G. M. 1990 Dynamics and controls of methane oxidation in a Danish wetland sediment. *FEMS Microbiol. Ecol.* **74**, 309–323.
- King, G. M. 1992 Ecological aspects of methane oxidation, a key determinant of global methane dynamics. *Adv. Microb. Ecol.* **12**, 431–468.
- King, G. M. 1993 Ecophysiological characteristics of obligate methanotrophic bacteria and methane oxidation *in situ*. In *Microbial growth on C1 compounds* (ed. J. C. Murrell & D. P. Kelly), pp. 303–313. Andover: Intercept.
- Knowles, R. 1982 Denitrification. *Microbiol. Rev.* **46**, 43–70.
- Knowles, R. & Topp, E. 1988 Some factors affecting nitrification and the production of nitrous oxide by the methanotrophic bacterium *Methylosinus trichosporium* OB3b. In *Current perspectives in environmental biogeochemistry* (ed. G. Giovannozzi-Sermanni & P. Nannipieri), pp. 383–393. Roma: CNR-IPRA.

- Koschorreck, M. & Conrad, R. 1993 Oxidation of atmospheric methane in soil: measurements in the field, in soil cores and in soil samples. *Global Biogeochem. Cycles* **7**, 109–121.
- Krämer, M., Baumgärtner, M., Bender, M. & Conrad, R. 1990 Consumption of NO by methanotrophic bacteria in pure culture and in soil. *FEMS Microbiol. Ecol.* **73**, 345–350.
- LaFavre, J. S. & Focht, D. D. 1983 Conservation in soil of H<sub>2</sub> liberated from N<sub>2</sub> fixation by lup-nodules. *Appl. Environ. Microbiol.* **46**, 304–311.
- Mayer, H. P. & Conrad, R. 1990 Factors influencing the population of methanogenic bacteria and the initiation of methane production upon flooding of paddy soil. *FEMS Microbiol. Ecol.* **73**, 103–112.
- Minami, K. & Fukushi, S. 1981 Volatilization of carbonyl sulphide from paddy soils treated with sulphur-containing substances. *Soil Sci. Plant Nutr.* **27**, 339–345.
- Möller, D. 1984 On the global natural sulphur emission. *Atmos. Environ.* **18**, 29–39.
- Mörsdorf, G., Frunzke, K., Gadkari, D. & Meyer, O. 1992 Microbial growth on carbon monoxide. *Biodeg.* **3**, 61–82.
- Novelli, P. C., Masarie, K. A., Tans, P. P. & Lang, P. M. 1994 Recent changes in atmospheric carbon monoxide. *Science, Wash.* **263**, 1587–1590.
- Peters, V. & Conrad, R. 1995 Methanogenic and other strictly anaerobic bacteria in desert soil and other oxic soils. *Appl. environ. Microbiol.* (In the press.)
- Popelier, F., Liessens, J. & Verstraete, W. 1985 Soil H<sub>2</sub>-uptake in relation to soil properties and rhizobial H<sub>2</sub>-production. *Plant Soil* **85**, 85–96.
- Prinn, R. G. 1994 Global atmospheric–biospheric chemistry. In *Global atmospheric–biospheric chemistry* (ed. R. G. Prinn), pp. 1–18. New York: Plenum Press.
- Protoschill-Krebs, G. & Kesselmeier, J. 1992 Enzymatic pathways for the consumption of carbonyl sulphide (COS) by higher plants. *Bot. Acta* **105**, 206–212.
- Reeburgh, W. S., Whalen, S. C. & Alperin, M. J. 1993 The role of methylotrophy in the global methane budget. In *Microbial growth on C1 compounds* (ed. J. C. Murrell & D. P. Kelly), pp. 1–14. Andover: Intercept.
- Remde, A. & Conrad, R. 1991 Metabolism of nitric oxide in soil and denitrifying bacteria. *FEMS Microbiol. Ecol.* **85**, 81–93.
- Remde, A., Slemr, F. & Conrad, R. 1989 Microbial production and uptake of nitric oxide in soil. *FEMS Microbiol. Ecol.* **62**, 221–230.
- Scharffe, D., Hao, W. M., Donoso, L., Crutzen, P. J. & Sanhueza, E. 1990 Soil fluxes and atmospheric concentration of CO and CH<sub>4</sub> in the northern part of the Guayana Shield, Venezuela. *J. geophys. Res.* **95**, 22475–22480.
- Schütz, H., Conrad, R., Goodwin, S. & Seiler, W. 1988 Emission of hydrogen from deep and shallow freshwater environments. *Biogeochem.* **5**, 295–311.
- Schütz, H., Schröder, P. & Rennenberg, H. 1991 Role of plants in regulating the methane flux to the atmosphere. In *Trace gas emissions by plants* (ed. T. D. Sharkey, E. A. Holland & H. A. Mooney), pp. 29–63. San Diego: Academic Press.
- Schuler, S. & Conrad, R. 1990 Soils contain two different activities for oxidation of hydrogen. *FEMS Microbiol. Ecol.* **73**, 77–84.
- Schuler, S. & Conrad, R. 1991 Hydrogen oxidation in soil following rhizobial H<sub>2</sub> production due to N<sub>2</sub> fixation by a *Vicia faba*-*Rhizobium leguminosarum* symbiosis. *Biol. Fertil. Soils* **11**, 190–195.
- Seiler, W. 1978 The influence of the biosphere on the atmospheric CO and H<sub>2</sub> cycles. In *Environmental biogeochemistry and geomicrobiology*, vol. 3 (ed. W. Krumbein), pp. 773–810. Ann Arbor, MI: Ann Arbor Science Publ.
- Seiler, W. & Conrad, R. 1981 Field measurements of natural and fertilizer induced N<sub>2</sub>O release rates from soils. *J. Air Poll. Contr. Ass.* **31**, 767–772.
- Sexstone, A. J., Revsbech, N. P., Parkin, T. B. & Tiedje, J. M. 1985 Direct measurement of oxygen profiles and denitrification rates in soil aggregates. *Soil Sci. Soc. Am. J.* **49**, 645–651.

- Sharkey, T. D., Holland, E. A., Mooney, H. A. (eds) 1991 *Trace gas emissions by plants*. San Diego: Academic Press.
- Slemr, F. & Seiler, W. 1984 Field measurements of NO and NO<sub>2</sub> emissions from fertilized and unfertilized soils. *J. atmos. Chem.* **2**, 1–24.
- Slemr, F. & Seiler, W. 1991 Field study of environmental variables controlling the NO and NO<sub>2</sub> emissions from soil, and of the NO and NO<sub>2</sub> compensation points. *J. geophys. Res.* **96**, 13017–13031.
- Steele, L. P., Dlugokencky, E. J., Lang, P. M., Tans, P. P., Martin, R. C. & Masarie, K. A. 1992 Slowing down of the global accumulation of atmospheric methane during the 1980s. *Nature, Lond.* **358**, 313–316.
- Tiedje, J. M. 1988 Ecology of denitrification and dissimilatory nitrate reduction to ammonia. In *Biology of anaerobic microorganisms* (ed. A. J. B. Zehnder), pp. 179–244. New York: Wiley.
- Yoshinari, T. 1985 Nitrite and nitrous oxide production by *Methylosinus trichosporium*. *Can. J. Microbiol.* **31**, 139–144.
- Zafriou, O. C., Hanley, Q. S. & Snyder, G. 1989 Nitric oxide and nitrous oxide reduction and cycling during dissimilatory nitrite reduction by *Pseudomonas perfectomarina*. *J. Biol. Chem.* **264**, 5694–5699.
- Zausig, J., Stepniewski, W. & Horn, R. 1993 Oxygen concentration and redox potential gradients in unsaturated model soil aggregates. *Soil Sci. Soc. Am. J.* **57**, 908–916.

#### Discussion

K. A. SMITH (*SAC, Edinburgh, U.K.*). In the course of work done in the U.S.A. over 20 years ago (Smith *et al.* 1973), which was primarily intended to study soil uptake of CO, I found that sterilized soils released CO, so presumably this was by a non-biological process. Does Dr Conrad believe that his observations of CO emission from soils in hot, dry conditions were due to non-biological production?

R. CONRAD. Yes, I can confirm that Dr Smith's observation that CO production in soil is largely due to non-biological processes that are also active in sterilized soil. CO is probably produced by thermal decomposition of humic material (Conrad & Seiler 1985a).

#### Additional references

- Smith, K. A., Brenner, J. M. & Tabatabai, M. A. 1973 Sorption of gaseous atmospheric pollutants by soils. *Soil Sci.* **116**, 313–319.